

# OptiPrep™ Mini-Review MV05

## Purification and analysis of papillomaviruses

- ◆ OptiPrep™ is a sterile 60% (w/v) solution of iodixanol in water, density = 1.32 g/ml
- ◆ This Mini-Review principally provides (in Section 2) a bibliography of all those papers reporting the use of OptiPrep™ in the purification and analysis of papilloma virus and virus-like particles. Section 1 briefly summarizes the advantages of using OptiPrep™; the gradient strategy used for papillomavirus and the technical data that is available.

### 1. Technical background to the use of OptiPrep™

#### 1a. Background

In all comparative studies between CsCl and iodixanol, the recovery of virus infectivity is much higher and the particle:infectivity ratio much lower when viruses are purified in iodixanol. Although sucrose is generally less deleterious to viral infectivity than CsCl, it can nevertheless also have serious effects on certain important aspects of viral function; in particular the loss of surface glycoproteins from enveloped viruses, particularly retroviruses, has been noted [1]. This may be related to its viscosity, which, in solutions of the same density, is much higher than that of iodixanol. Most iodixanol gradients can also be made isoosmotic over the entire density range.

Like CsCl, sucrose must be dialyzed before infectivity can be measured. In contrast both infectivity measurements using cultured cells and many add-on techniques can be performed without dialysis of iodixanol. Combined with the availability of OptiPrep™ as a sterile solution, this makes the use of OptiPrep™ for virus purification and assembly analysis much more convenient than the use of either CsCl or sucrose. The only analytical technique for which removal of the iodixanol is essential is electron microscopy. Consequently iodixanol is being increasingly used for the purification of papillomavirus particles from lysed cultured cells.

#### 1b. Gradient strategy

Buck et al [2] observed that papillomavirus vector stocks could be purified by an iodixanol gradient centrifugation procedure that was substantially more effective than the standard CsCl gradient purification. The methodology developed [2] has been used more or less unchanged in all subsequent studies and has been found to be effective for human, bovine and cotton-tail rabbit viruses. A continuous gradient is usually generated from equal volumes of 27%, 33% and 39% (w/v) iodixanol allowed to diffuse for 3-4 h at room temperature, after which the clarified sample is laid on top. Although the continuous gradient might be prepared from 27% and 39% iodixanol using a two-chamber gradient maker, use of the latter is technically more difficult with these small volume gradients. The separation of the L1 protein from both DNA-containing and empty virus-like particles after 234,000 g for 3.5 h at 16°C is considered to be part buoyant density and part sedimentation velocity [2], so top-loading of the sample cannot be replaced by other frequently-used strategies such as bottom-loading of the sample or the use of self-generated gradients.

#### 1c. References (to Sections 1a and 1b)

1. Palker, T.J. (1990) *Mapping of epitopes on human T-cell leukemia virus type 1 envelope glycoprotein* In: Human Retrovirology: HTLV (ed. Blattner, W.A.) Raven Press, NY, pp 435-445
2. Buck, C.B., Pastrana, D.V., Lowy, D.R. and Schiller, J.T. (2004) *Efficient intracellular assembly of papillomaviral vectors* J. Virol., **78**, 751-757

- ◆ The gradient protocol for the purification of isolation of papillomavirus particles (Application Sheet V10) may be accessed from the OptiPrep™ Applications flash-drive or from the following website: [www.axis-shield-density-gradient-media.com](http://www.axis-shield-density-gradient-media.com), click on “Methodology” then “Viruses” to open up the Virus Index. Other OptiPrep™ Application Sheets on the preparation and harvesting of gradients may also be accessed from the top of the Index.

## 2. Bibliography

- ◆ This bibliography provides a comprehensive reference list of all the papers reporting the use of OptiPrep™ for papillomavirus purification, published before the end of January 2017.
- ◆ The references are divided alphabetically into “**Research topic**” sections and subsections.
- ◆ All references are listed alphabetically according to **First Author**.
- ◆ The vast majority of published papers describe studies on human papillomavirus (not flagged); those studies on bovine (**B**), canine (**C**), cotton-tailed rabbit (**CTR**), equine (**E**), Macaques (**MQ**) and mouse (**M**) are indicated as shown. In many cases these flagged papers will also include work on the human virus.

### B-cell anergy reversal

**Chackerian, B.**, Durfee, M.R. and Schiller, J.T. (2008) *Virus-like display of a neo-self antigen reverses B cell anergy in a B cell receptor transgenic mouse model* J. Immunol., **180**, 5816-5825 (**B**)

### Baculoviral vectors

**Cho, H.**, Lee, H-J., Heo, Y-K., Cho, Y., Gwon, Y-D., Kim, M-G., Park, K.H., Oh, Y-K. and Kim, Y.B. (2014) *Immunogenicity of a trivalent human papillomavirus L1 DNA-encapsidated, non-replicable baculovirus nanovaccine* PLoS One, **9**: e95961

**Lee, H-J.**, Hur, Y-K., Cho, Y-D., Kim, M-G., Lee, H-T., Oh, Y-K. and Kim, Y.B. (2012) *Immunogenicity of bivalent human papillomavirus DNA vaccine using human endogenous retrovirus envelope-coated baculoviral vectors in mice and pigs* PLoS One, **7**: e50296

**Lee, H-J.**, Cho, H., Kim, M-G., Heo, Y-K., Cho, Y., Gwon, Y-D., Park, K.H., Jin, H., Kim, J., Oh, Y-K. and Kim, Y.B. (2015) *Sublingual immunization of trivalent human papillomavirus DNA vaccine in baculovirus nanovector for protection against vaginal challenge* PLoS One, **10**: e0119408

### Capsids and capsid protein

#### Capsid assembly

**Day, P.M.**, Thompson, C.D., Pang, Y.Y., Lowy, D.R. and Schiller, J.T. (2015) *Involvement of nucleophosmin (NPM1/B23) in assembly of infectious HPV16 capsids* Papillomavirus Res. **1**, 74–89

#### DNA, separation from

**Bzhalava, D.**, Johansson, H., Ekström, J., Faust, H., Möller, B., Eklund, C., Nordin, P., Stenquist, B., Paoli, J., Persson, B., Forslund, O. and Dillner, J. (2013) *Unbiased approach for virus detection in skin lesions* PLoS One, **8**: e65953

#### Dynein interacting domains

**Florin, L.**, Becker, K.A., Lambert, C., Nowak, T., Sapp, C., Strand, D., Streeck, R.E. and Sapp, M. (2006) *Identification of a dynein interacting domain in the papillomavirus minor capsid protein L2* J. Virol., **80**, 6691-6696

#### L1 capsid protein

**Bienkowska-Haba, M.**, Williams, C., Kim, S.M., Garcea, R.L. and Sapp, M. (2012) *Cyclophilins facilitate dissociation of the human papillomavirus type 16 capsid protein L1 from the L2/DNA complex following virus entry* J. Virol., **86**, 9875-9887

**Ishii, Y.**, Kondo, K., Matsumoto, T., Tanaka, K., Shinkai-Ouchi, F., Hagiwara, K. and Kanda, T. (2007) *Thiol-reactive reagents inhibits intracellular trafficking of human papillomavirus type 16 pseudovirions by binding to cysteine residues of major capsid protein L1* Virol. J., **4**:110

**Mistry, N.**, Wibom, C. and Evander, M. (2008) *Cutaneous and mucosal human papillomaviruses differ in net surface charge, potential impact on tropism* Virol. J., **5**:118

**Ryndock, E.J.**, Conway, M.J., Alam, S., Gul, S., Murad, S., Christensen, N.D. and Meyers, C. (2014) *Roles for human papillomavirus type 16 L1 cysteine residues 161, 229, and 379 in genome encapsidation and capsid stability* PLoS One, **9**: e99488

#### Maturation

**Buck, C.B.**, Thompson, C.D., Pang, Y-Y-s., Lowy, D.R. and Schiller, J.T. (2005) *Maturation of papillomavirus capsids* J. Virol., **79**, 2839-2846 (**B**)

**Cardone, G.**, Moyer, A.L., Cheng, N., Thompson, C.D., Dvoretzky, I., Lowy, D.R., Schiller, J.T., Steven, A.C., Buck, C.B. and Trus, B.L. (2014) *Maturation of the human papillomavirus 16 capsid* mBio, **5**: e01104-14

**Conway, M.J.**, Cruz, L., Alam, S., Christensen, N.D. and Meyers, C. (2011) *Differentiation-dependent interpentameric disulfide bond stabilizes native human papillomavirus type 16* PLoS One, **6**: e22427

### Neutralization-sensitive epitopes

**Culp, T.D.**, Spatz, C.M., Reed, C.A. and Christensen, N.D. (2007) *Binding and neutralization efficiencies of monoclonal antibodies, Fab fragments and scFv specific for L1 epitopes on the capsid of infectious HPV particles* Virology, **361**, 435-446

### Tumour cell binding

**Kines, R.C.**, Cerio, R.J., Roberts, J.N., Thompson, C.D., de Los Pinos, E., Lowy, D.R. and Schiller, J.T. (2016) *Human papillomavirus capsids preferentially bind and infect tumor cells* Int. J. Cancer, **138**, 901–911

### Cell entry/targeting

#### Autophagy inhibition

**Surviladze, Z.**, Sterk, R.T., DeHaro, S.A. and Ozbun, M.A. (2013) *Cellular entry of human papillomavirus type 16 involves activation of the phosphatidylinositol 3-kinase/Akt/mTOR pathway and inhibition of autophagy* J. Virol., **87**, 2508–2517

#### Clathrin/caveolin

**Spoden, G.**, Freitag, K., Husmann, M., Boller, K., Sapp, M., Lambert, C. and Florin, L. (2008) *Clathrin- and caveolin-independent entry of human papillomavirus type 16 - involvement of tetraspanin-enriched microdomains (TEMs)* PLoS One, **3**:e3313

#### Cyclophilin receptors

**Bienkowska-Haba, M.**, Patel, H.D. and Sapp, M. (2009) *Target cell cyclophilins facilitate human papillomavirus type 16 infection* PLoS Pathog., **5**:e1000524 (B)

#### Cysteine proteases

**Dabydeen, S.A.** and Meneses, P.I. (2009) *The role of NH<sub>4</sub>Cl and cysteine proteases in human papillomavirus type 16 infection* Virol. J., **6**:109

#### Dynamin inhibition

**Abban, C.Y.**, Bradbury, N.A. and Meneses, P.I. (2008) *HPV16 and BPV1 infection can be blocked by the dynamin inhibitor dynasore* Am. J. Therapeut., **15**, 304-311 (B)

#### Dynein light chain requirement

**Schneider, M.A.**, Spoden, G.A., Florin, L. and Lambert, C. (2011) *Identification of the dynein light chains required for human papillomavirus infection* Cell. Microbiol., **13**, 32–46

#### Focal adhesion kinase activation

**Abban, C.Y.** and Meneses, P.I. (2010) *Usage of heparan sulfate, integrins, and FAK in HPV16 infection* Virology **403**, 1–16

#### Heparan sulphate receptors

**Abban, C.Y.** and Meneses, P.I. (2010) *Usage of heparan sulfate, integrins, and FAK in HPV16 infection* Virology **403**, 1–16

**Day, P.M.**, Lowy, D.R. and Schiller, J.T. (2008) *Heparan sulfate-independent cell binding and infection with furin-precleaved papillomavirus capsids* J. Virol., **82**, 12565-12568

**Donalisio, M.**, Rusnati, M., Civra, A., Bugatti, A., Allemand, D., Pirri, G., Giuliani, A., Landolfo, S. and Lembo, D. (2010) *Identification of a dendrimeric heparan sulfate-binding peptide that inhibits infectivity of genital types of human papillomaviruses* Antimicrob. Agents Chemother., **54**, 4290-4299

**Johnson, K.M.**, Kines, R.C., Roberts, J.N., Lowy, D.R., Schiller, J.T. and Day, P.M. (2009) *Role of heparan sulfate in attachment to and infection of the murine female genital tract by human papillomavirus* J. Virol., **83**, 2067-2074

**Knappe, M.**, Bodevin, S., Selinka, H-C., Spillman, D., Streeck, R.E., Chen, X.S., Lindahl, U. and Sapp, M. (2007) *Surface-exposed amino acid residues of HPV16L1 protein mediating interaction with cell surface heparan sulfate* J. Biol. Chem., **282**, 27913-27922

**Selinka, H-C.**, Florin, L., Patel, H.D., Freitag, K., Schmidtke, M., Makarov, V.A. and Sapp, M. (2007) *Inhibition of transfer to secondary receptors by heparin sulfate-binding drug or antibody induces noninfectious uptake of human papillomavirus* J. Virol., **81**, 10970-10980

### **Inhibition by thio-reactive agents**

**Ishii, Y.**, Kondo, K., Matsumoto, T., Tanaka, K., Shinkai-Ouchi, F., Hagiwara, K. and Kanda, T. (2007) *Thiol-reactive reagents inhibits intracellular trafficking of human papillomavirus type 16 pseudovirions by binding to cysteine residues of major capsid protein L1* Virol. J., **4**:110

### **Laminin-5 binding**

**Culp, T.D.**, Budgeon, L.R., Marinkovich, M.P., Meneguzzi, G. and Christensen, N.D. (2006) *Keratinocyte-secreted laminin 5 can function as a transient receptor for human papillomaviruses by binding virions and transferring them to adjacent cells* J. Virol., **80**, 8940-8950

### **L1/L2 protein interactions**

**Buck, C.B.**, Cheng, N., Thompson, C.D., Lowy, D.R., Steven, A.C., Schiller, J.T. and Trus, B.L. (2008) *Arrangement of L2 within the papillomavirus capsid* J. Virol., **82**, 5190-5197

**Knappe, M.**, Bodevin, S., Selinka, H-C., Spillman, D., Streeck, R.E., Chen, X.S., Lindahl, U. and Sapp, M. (2007) *Surface-exposed amino acid residues of HPV16L1 protein mediating interaction with cell surface heparan sulfate* J. Biol. Chem., **282**, 27913-27922

### **Microtubules, L2 interaction with**

**Schneider, M.A.**, Spoden, G.A., Florin, L. and Lambert, C. (2011) *Identification of the dynein light chains required for human papillomavirus infection* Cell. Microbiol., **13**, 32–46

### **Phosphatidylinositol-3 kinase**

**Surviladze, Z.**, Sterk, R.T., DeHaro, S.A. and Ozbun, M.A. (2013) *Cellular entry of human papillomavirus type 16 involves activation of the phosphatidylinositol 3-kinase/Akt/mTOR pathway and inhibition of autophagy* J. Virol., **87**, 2508–2517

### **PML expression**

**Day, P.M.**, Baker, C.C., Lowy, D.R. and Schiller, J.T. (2004) *Establishment of papillomavirus infection is enhanced by promyelocytic leukemia protein (PML) expression* Proc. Natl. Acad. Sci. USA, **101**, 14252-14257 **(B)**

### **γ-Secretase requirement**

**Huang, H-S.**, Buck, C.B. and Lambert, P.F. (2010) *Inhibition of gamma secretase blocks HPV infection* Virology **407**, 391–396

**Karanam, B.**, Peng, S., Li, T., Buck, C., Day, P.M. and Roden, R.B.S. (2010) *Papillomavirus infection requires γ secretase* J. Virol., **84**, 10661–10670 **(CTR)**

### **Tetraspannin-enriched domains**

**Scheffer, K.D.**, Gawlitza, A., Spoden, G.A., Zhang, X.A., Lambert, C., Berditchevski, F. and Florina, L. (2013) *Tetraspanin CD151 mediates papillomavirus type 16 endocytosis* J. Virol., **87**, 3435–3446

**Spoden, G.**, Freitag, K., Husmann, M., Boller, K., Sapp, M., Lambert, C. and Florin, L. (2008) *Clathrin- and caveolin-independent entry of human papillomavirus type 16 - involvement of tetraspanin-enriched microdomains (TEMs)* PLoS One, **3**:e3313

### **Epithelial cell, expression in**

**Israr, M.**, Biryukov, J., Ryndock, E.J., Alam, S. and Meyers, C. (2016) *Comparison of human papilloma-virus type16 replication in tonsil and foreskin epithelia* Virology, **499**, 82–90

### **Gene expression**

**Berg, M.**, Gambhira, R., Siracusa, M., Hoiczky, E., Roden, R. and Ketner, G. (2007) *HPV16L1 capsid protein expressed from viable adenovirus recombinant elicits neutralizing antibody in mice* Vaccine, **25**, 3501-3510

### **Genome**

#### **Amplification**

**Culp, T.D.**, Cladel, N.M., Balogh, K.K., Budgeon, L.R., Mejia, A.F. and Christensen, N.D. (2006) *Papillomavirus particles assembled in 293TT cells are infectious in vivo* J. Virol., **80**, 11381-11384 **(CTR)**

#### **Encapsulation**

**Pyeon, D.**, Lambert, P.F. and Ahlquist, P. (2005) *Production of infectious human papillomavirus independently of viral replication and epithelial cell differentiation* Proc. Natl. Acad. Sci. USA, **102**, 9311-9316

## Packaging

Cerqueira, C., Pang, Y.-Y.S., Day, P.M., Thompson, C.D., Buck, C.B., Lowy, D.R. and Schiller, J.T. (2016) *A cell-free assembly system for generating infectious human papillomavirus 16 capsids implicates a size discrimination mechanism for preferential viral genome packaging* J. Virol., **90**, 1096-1107

## Immunogenicity

**Kwag, H.-L.**, Kim, H.J., Chang, D.Y. and Kim, H.-J. (2012) *The production and immunogenicity of human papillomavirus type 58 virus-like particles produced in Saccharomyces cerevisiae* J. Microbiol., **50**, 813-820

**Lee, H.-J.**, Hur, Y.-K., Cho, Y.-D., Kim, M.-G., Lee, H.-T., Oh, Y.-K. and Kim, Y.B. (2012) *Immunogenicity of bivalent human papillomavirus DNA vaccine using human endogenous retrovirus envelope-coated baculoviral vectors in mice and pigs* PLoS One, **7**: e50296

## Infection

### Anti-L1 antibodies

**Hu, J.**, Budgeon, L.R., Cladel, N.M., Culp, T.A., Balogh, K.K. and Christensen, N.D. (2007) *Detection of L1, infectious virions and anti-L1 antibody in domestic rabbits infected with cottontail rabbit papillomavirus* J. Gen. Virol., **88**, 3286-3293 (CTR)

### Antivirals

**Huang, H.-S.**, Pyeon, D., Pearce, S.M., Lank, S.M., Griffin, L.M., Ahlquist, P., Lambert, P.F. (2012) *Novel antivirals inhibit early steps in HPV infection* Antiviral Res., **93**, 280-287

**Theisen, L.L.**, Erdelmeier, C.A.J., Spoden, G.A., Boukhallouk, F., Sausy, A., Florin, L. and Muller, C.P. (2014) *Tannins from Hamamelis virginiana bark extract: characterization and improvement of the antiviral efficacy against influenza A virus and human papillomavirus* PLoS One, **9**: e88062

### Cell cycle progression requirement

**Pyeon, D.**, Pearce, S.M., Lank, S.M., Ahlquist, P. and Lambert, P.F. (2009) *Establishment of human papillomavirus infection requires cell cycle progression* PLoS Pathog., **5**: e1000318

### L2 Cysteine residues

**Gambhira, R.**, Jagu, S., Karanam, B., Day, P.M. and Roden, R. (2009) *Role of L2 cysteines in papillomavirus infection and neutralization* Virol. J., **6**: 176 (B)

### Heparan sulphate

**Cagno, V.**, Donalisio, M., Bugatti, A., Civra, A., Cavalli, R., Ranucci, E., Ferruti, P., Rusnati, M. and Lembo, D. (2015) *The agmatine-containing poly(amidoamine) polymer AGMA1 binds cell surface heparan sulfates and prevents attachment of mucosal human papillomaviruses* Antimicrob. Agents Chemother., **59**, 5250-5259

**Huang, H.-S.** and Lambert, P.F. (2012) *Use of an in vivo animal model for assessing the role of integrin  $\alpha_6\beta_4$  and Syndecan-1 in early steps in papillomavirus infection* Virology, **433**, 395-400

**Johnson, K.M.**, Kines, R.C., Roberts, J.N., Lowy, D.R., Schiller, J.T. and Day, P.M. (2009) *Role of heparan sulfate in attachment to and infection of the murine female genital tract by human papillomavirus* J. Virol., **83**, 2067-2074

**Kines, R.C.**, Cerio, R.J., Roberts, J.N., Thompson, C.D., de Los Pinos, E., Lowy, D.R. and Schiller, J.T. (2016) *Human papillomavirus capsids preferentially bind and infect tumor cells* Int. J. Cancer, **138**, 901-911

**Kumar, A.**, Jacob, T., Abban, C.Y. and Meneses, P.I. (2014) *Intermediate heparan sulfate binding during HPV-16 infection in HaCaTs* Am. J. Therapeut., **21**, 331-342

**Lembo, D.**, Donalisio, M., Laine, C., Cagno, V., Civra, A., Bianchini, E.P., Zeghib, N. and Bouchemal, K. (2014) *Auto-associative heparin nanoassemblies: A biomimetic platform against the heparan sulfate-dependent viruses HSV-1, HSV-2, HPV-16 and RSV* Eur. J. Pharm. Biopharm., **88**, 275-282

### Integrins

**Aksoy, P.**, Abban, C.Y., Kiyashka, E., Qiang, W. and Meneses, P.I. (2014) *HPV16 infection of HaCaTs is dependent on  $\beta_4$  integrin, and  $\alpha_6$  integrin processing* Virology, **449**, 45-52

**Huang, H.-S.** and Lambert, P.F. (2012) *Use of an in vivo animal model for assessing the role of integrin  $\alpha_6\beta_4$  and Syndecan-1 in early steps in papillomavirus infection* Virology, **433**, 395-400

### Optical imaging

**Kines, R.C.**, Kobayashi, H., Choyke, P.L. and Bernardo, M.L. (2013) *Optical imaging of HPV infection in a murine model* In Mol. Dermatol: Methods and Protocols (ed. Has, C. and Sitaru, C.) Springer Science+Business Media, LLC, pp 141-150

### Restriction factors

**Warren, C.J.**, Xu, T., Guo, K., Griffin, L.M., Westrich, J.A., Lee, D., Lambert, P.F., Santiago, M.L. and Pyeona, D. (2015) *APOBEC3A functions as a restriction factor of human papillomavirus* J. Virol., 89, 688-702

### $\gamma$ -Secretase requirement

**Huang, H-S.**, Buck, C.B. and Lambert, P.F. (2010) *Inhibition of gamma secretase blocks HPV infection* Virology **407**, 391–396

**Karanam, B.**, Peng, S., Li, T., Buck, C., Day, P.M. and Roden, R.B.S. (2010) *Papillomavirus infection requires  $\gamma$  secretase* J. Virol., **84**, 10661–10670 (CTR)

### Single particle tracking

Ewers, H. and Schelhaas, M. (2012) *Analysis of virus entry and cellular membrane dynamics by single particle tracking* Methods Enzymol., **506**, 63-80

### Skin/vaginal lesions

**Bzhalava, D.**, Johansson, H., Ekström, J., Faust, H., Möller, B., Eklund, C., Nordin, P., Stenquist, B., Paoli, J., Persson, B., Forslund, O. and Dillner, J. (2013) *Unbiased approach for virus detection in skin lesions* PLoS One, **8**: e65953

**Chu, T-Y.**, Chang, Y-C. and Ding, D.C. (2013) *Cervicovaginal secretions protect from human papillomavirus infection: Effects of vaginal douching* Taiwan. J. Obstet. Gynecol., **52**, 241-245

**Çuburu, N.**, Cerio, R.J., Thompson, C.D. and Day, P.M. (2015) *Mouse model of cervicovaginal papillomavirus infection* In Cervical Cancer: Methods and Protocols, Methods in Molecular Biology, vol. 1249 (eds. Keppler, D. and Lin, A.W. Springer Science+Business Media New York, pp 365-379

**Handisurya, A.**, Day, P.M., Thompson, C.D., Buck, C.B., Kwak, K., Roden, R.B.S., Lowy, D.R. and Schiller, J.T. (2012) *Murine skin and vaginal mucosa are similarly susceptible to infection by pseudovirions of different papillomavirus classifications and species* Virology, **433**, 385–394 (M)

**Roberts, J.N.**, Kines, R.C., Katki, H.A., Lowy, D.R. and Schiller, J.T. (2011) *Effect of pap smear collection and carrageenan on cervicovaginal human papillomavirus-16 infection in a rhesus macaque model* J. Natl. Cancer Inst., **103**, 737–743

**Vinzón, S.E.**, Braspenning-Wesch, I., Müller, M., Geissler, E.K., Nindl, I., Gröne, H-J., Schäfer, K. and Rösl, F. (2014) *Protective vaccination against papillomavirus-induced skin tumors under immuno-competent and immunosuppressive conditions: a preclinical study using a natural outbred animal model* PLoS Pathog., **10**: e1003924 (M)

### Syndecan-1

**Huang, H-S.** and Lambert, P.F. (2012) *Use of an in vivo animal model for assessing the role of integrin  $\alpha_6\beta_4$  and Syndecan-1 in early steps in papillomavirus infection* Virology, **433**, 395–400

### Infection inhibition

#### Carrageenens

**Buck, C.B.**, Thompson, C.D., Roberts, J.N., Müller, M., Lowy, D.R. and Schiller, J.T. (2007) *Carrageenan is a potent inhibitor of papillomavirus function* Plos Pathog., **2**:e69

**Novetsky, A.P.**, Keller, M.J., Gradissimo, A., Chen, Z., Morgan, S.L., Xue, X., Strickler, H.D., Fernández-Romero, J.A., Burk, R. and Einstein, M.H. (2016) *In vitro inhibition of human papillomavirus following use of a carrageenan-containing vaginal gel* Gynecol. Oncol., **143**, 313–318

### Cholesterol derivatives

**Civra, A.**, Cagno, V., Donalisio, M., Biasi, F., Leonarduzzi, G., Poli, G. and Lembo, D. (2014) *Inhibition of pathogenic non-enveloped viruses by 25-hydroxycholesterol and 27-hydroxycholesterol* Sci. Rep., **4**: 7487

### $\alpha$ -Defensins

**Buck, C.B.**, Day, P.M., Thompson, C.D., Lubkowski, J., Lu, W., Lowy, D.R. and Schiller, J.T. (2006) *Human  $\alpha$ -defensins block papillomavirus infection* Proc. Natl. Acad. Sci. USA, **103**, 1516-1521

**Gounder, A.P.**, Wiens, M.E., Wilson, S.S., Lu, W. and Smith, J.G. (2012) *Critical determinants of human  $\alpha$ -defensin 5 activity against non-enveloped viruses* J. Biol. Chem., **287**, 24554–24562

**Wiens, M.E.** and Smith, J.G. (2015) *Alpha-defensin HD5 inhibits furin cleavage of human papillomavirus 16 L2 to block infection* J. Virol., **89**, 2866-2874

### **E. coli sulphated polysaccharides**

**Lembo, D.**, Donalisio, M., Rusnati, M., Bugatti, A., Cornaglia, M., Cappello, P., Giovarelli, M., Oreste, P. and Landolfo, S. (2008) *Sulfated K5 Escherichia coli polysaccharide derivatives as wide-range inhibitors of genital types of human papillomavirus* Antimicrob. Agents Chemother., **52**, 1374-1381

### **Genome, nuclear transport block**

**Ishii, Y.**, Tanaka, K., Kondo, K., Takeuchi, T., Mori, S. and Kanda, T. (2010) *Inhibition of nuclear entry of HPV16 pseudovirus-packaged DNA by an anti-HPV16 L2 neutralizing antibody* Virology **406**, 181–188

### **Heparan sulphate binding**

**Cagno, V.**, Donalisio, M., Bugatti, A., Civra, A., Cavalli, R., Ranucci, E., Ferruti, P., Rusnati, M. and Lembo, D. (2015) *The agmatine-containing poly(amidoamine) polymer AGMA1 binds cell surface heparan sulfates and prevents attachment of mucosal human papillomaviruses* Antimicrob. Agents Chemother., **59**, 5250-5259

**Donalisio, M.**, Rusnati, M., Civra, A., Bugatti, A., Allemand, D., Pirri, G., Giuliani, A., Landolfo, S. and Lembo, D. (2010) *Identification of a dendrimeric heparan sulfate-binding peptide that inhibits infectivity of genital types of human papillomaviruses* Antimicrob. Agents Chemother., **54**, 4290-4299

### **High affinity ligands**

**Mauro, N.**, Ferruti, P., Ranucci, E., Manfredi, A., Berzi, A., Clerici, M., Cagno, V., Lembo, D., Palmioli, A. and Sattin, S. (2016) *Linear biocompatible glycopolyamidoamines as dual action mode virus infection inhibitors with potential as broad-spectrum microbicides for sexually transmitted diseases* Sci. Rep., **6**: 33393

### **L1/L2 antibody, comparison of**

**Day, P.M.**, Kines, R.C., Thompson, C.D., Jagu, S., Roden, R.B., Lowy, D.R. and Schiller, J.T. (2010) *In vivo mechanisms of vaccine-induced protection against HPV infection* Cell Host Microbe, **8**, 260–270

### **Mucins**

**Lieleg, O.**, Lieleg, C., Bloom, J., Buck, C.B. and Ribbeck, K. (2012) *Mucin biopolymers as broad-spectrum antiviral agents* Biomacromolecules, **13**, 1724–1732

### **Infectious virus production**

#### **Genome incorporation**

**Pyeon, D.**, Lambert, P.F. and Ahlquist, P. (2005) *Production of infectious human papillomavirus independently of viral replication and epithelial cell differentiation* Proc. Natl. Acad. Sci. USA, **102**, 9311-9316

#### **Vacuolar ATPase**

**Müller, K.H.**, Spoden, G.A., Scheffer, K.D., Brunnhöfer, R., De Brabander, J.K., Maier, M.E., Florin, L. and Muller, C.P. (2014) *Inhibition by cellular vacuolar ATPase impairs human papillomavirus uncoating and infection* Antimicrob. Agents Chemother., **58**, 2905–2911

### **Intracellular assembly**

#### **L1/L2 proteins**

**Bissa, M.**, Zanutto, C., Pacchioni, S., Volonté, L., Venuti, A., Lembo, D., De Giulio Morghen, C. and Radaelli, A. (2015) *The L1 protein of human papilloma virus 16 expressed by a fowlpox virus recombinant can assemble into virus-like particles in mammalian cell lines but elicits a non-neutralising humoral response* Antiviral Res., **116**, 67–75

**Buck, C.B.**, Pastrana, D.V., Lowy, D.R. and Schiller, J.T. (2004) *Efficient intracellular assembly of Papillomaviral vectors* J. Virol., **78**, 751-757 (**B**)

**Buck, C.B.**, Cheng, N., Thompson, C.D., Lowy, D.R., Steven, A.C., Schiller, J.T. and Trus, B.L. (2008) *Arrangement of L2 within the papillomavirus capsid* J. Virol., **82**, 5190-5197

**Conway, M.J.** and Meyers, C. (2009) *Replication and assembly of human papillomaviruses* Dent. Res., **88**, 307-317 (**CTR**)

#### **L2 cysteine residues**

**Conway, M.J.**, Alam, S., Christensen, N.D. and Meyers, C. (2009) *Overlapping and independent structural roles for human papillomavirus type 16 L2 conserved cysteines* Virology **393**, 295–303

**Gambhira, R.**, Jagu, S., Karanam, B., Day, P.M. and Roden, R. (2009) *Role of L2 cysteines in papillomavirus infection and neutralization* Virol. J., **6**: 176 (**B**)

### Redox gradient dependence

**Conway, M.J.**, Alam, S., Ryndock, E.J., Cruz, L., Christensen, N.D., Roden, R.B.S. and Meyers, C. (2009) *Tissue-spanning redox gradient-dependent assembly of native human papillomavirus type 16 virions* J. Virol., **83**, 10515-10526

### Intracellular trafficking:

#### Cysteine proteases

**Dabydeen, S.A.** and Meneses, P.I. (2009) *The role of NH<sub>4</sub>Cl and cysteine proteases in human papillomavirus type 16 infection* Virol. J., **6**:109

#### Dynein interacting domain

**Florin, L.**, Becker, K.A., Lambert, C., Nowak, T., Sapp, C., Strand, D., Streeck, R.E. and Sapp, M. (2006) *Identification of a dynein interacting domain in the papillomavirus minor capsid protein L2* J. Virol., **80**, 6691-6696

### Endosomal trafficking

**Gräbel, L.**, Fast, L.A., Scheffer, K.D., Boukhallouk, F., Spoden, G.A., Tenzer, S., Boller, K., Bago, R., Rajesh, S. et al (2016) *The CD63-syntenin-1 complex controls post-endocytic trafficking of oncogenic human papillomaviruses* Sci. Rep., **6**: 32337

**Popa, A.**, Zhang, W., Harrison, M.S., Goodner, K., Kazakov, T., Goodwin, E.C., Lipovsky, A., Burd, C.G. and DiMaio, D. (2015) *Direct binding of retromer to human papillomavirus type 16 minor capsid protein L2 mediates endosome exit during viral infection* PLoS Pathog., **11**: e1004699

### Genome, nuclear transport block

**Ishii, Y.**, Tanaka, K., Kondo, K., Takeuchi, T., Mori, S. and Kanda, T. (2010) *Inhibition of nuclear entry of HPV16 pseudovirus-packaged DNA by an anti-HPV16 L2 neutralizing antibody* Virology **406**, 181–188

### Inhibition by thio-reactive agents

**Ishii, Y.**, Kondo, K., Matsumoto, T., Tanaka, K., Shinkai-Ouchi, F., Hagiwara, K. and Kanda, T. (2007) *Thiol-reactive reagents inhibits intracellular trafficking of human papillomavirus type 16 pseudovirions by binding to cysteine residues of major capsid protein L1* Virol. J., **4**:110

### L2 capsid protein

**Kondo, K.**, Ishii, Y., Mori, S., Shimabukuro, S., Yoshikawa, H. and Kanda, T. (2009) *Nuclear location of minor capsid protein L2 is required for expression of a reporter plasmid packaged in HPV51 pseudovirions* Virology **394**, 259–265

**Mamoor, S.**, Onder, Z., Karanam, B., Kwak, K., Bordeaux, J., Crosby, L., Roden, R.B.S. and Moroianu, J. (2012) *The high risk HPV16 L2 minor capsid protein has multiple transport signals that mediate its nucleocytoplasmic traffic* Virology, **422**, 413–424

### PML expression

**Day, P.M.**, Baker, C.C., Lowy, D.R. and Schiller, J.T. (2004) *Establishment of papillomavirus infection is enhanced by promyelocytic leukemia protein (PML) expression* Proc. Natl. Acad. Sci. USA, **101**, 14252-14257 **(B)**

### Polyethylenimines

**Spoden, G.A.**, Besold, K., Krauter, S., Plachter, B., Hanik, N., Kilbinger, A.F.M., Lambert, C. and Florina, L. (2012) *Polyethylenimine is a strong inhibitor of human papillomavirus and cytomegalovirus infection* Antimicrob. Agents Chemother., **56**, 75-82

### Keratinocyte/expression in/interactions

**Brendle, S.A.** and Christensen, N.D. (2015) *HPV binding assay to Laminin-332/Integrin  $\alpha 6\beta 4$  on human keratinocytes* In Cervical Cancer: Methods and Protocols, Methods in Molecular Biology, vol. 1249 (eds. Keppler, D. and Lin, A.W. Springer Science+Business Media New York, pp 53-66

**McKinney, C.C.**, Kim, M.J., Chen, D. and McBride, A.A. (2016) *Brd4 activates early viral transcription upon human papillomavirus 18 infection of primary keratinocytes* mBio, **7**: e01644-16

**Tao, L.**, Pavlova, S.I., Gasparovich, S.R., Jin, L. and Schwartz, J. (2015) *Alcohol metabolism by oral Streptococci and interaction with human papillomavirus leads to malignant transformation of oral keratinocytes* In Advances in Experimental Medicine and Biology, **815** Biological Basis of Alcohol-Induced Cancer. (ed. Vasiliou, V. et al), Springer International Publishing Switzerland pp 239-264

**Van Doorslaer, K.**, Porter, S., McKinney, C., Stepp, W.H. and McBride, A.A. (2016) *Novel recombinant papillomavirus genomes expressing selectable genes* Sci. Rep., **6**: 37782

### Langerhans cell activation

**Da Silva, D.M.**, Movius, C.A., Raff, A.B., Brand, H.E., Skeate, J.G., Wong, M.K. and Kast, W.M. (2014) *Suppression of Langerhans cell activation is conserved amongst human papillomavirus  $\alpha$  and  $\beta$  genotypes, but not a  $\mu$  genotype* Virology, **452-453**, 279–286

### Methodology

**Buck, C.B.**, Pastrana, D.V., Lowy, D.R. and Schiller, J.T. (2004) *Efficient intracellular assembly of Papillomaviral vectors* J. Virol., **78**, 751-757 (**B**)

**Buck, C.B.**, Pastrana, D.V., Lowy, D.R. and Schiller, J.T. (2005) *Generation of HPV pseudovirions using transfection and their use in neutralization assays* Methods Mol. Med., **119**, 445-462

### Neutralization assays

**Buck, C.B.**, Pastrana, D.V., Lowy, D.R. and Schiller, J.T. (2005) *Generation of HPV pseudovirions using transfection and their use in neutralization assays* Methods Mol. Med., **119**, 445-462

**Lamprecht, R.L.**, Kennedy, P., Huddy, S.M., Bethke, S., Hendrikse, M., Hitzeroth, I.I. and Rybicki, E.P. (2016) *Production of human papillomavirus pseudovirions in plants and their use in pseudovirion-based neutralization assays in mammalian cells* Sci. Rep., **6**: 20431

**Pastrana, D.V.**, Buck, C.B., Pang, Y-Y. S., Thompson, C.D., Castle, P.E., Fitzgerald, P.C., Kjaer, S.K., Lowy, D.R. and Schiller, J.T. (2004) *Reactivity of human sera in a sensitive, high-throughput pseudovirus-based papillomavirus neutralization assay for HPV16 and HPV18* Virology, **321**, 205-216

**Sehr, P.**, Rubio, I., Seitz, H., Putzker, K., Ribeiro-Müller, L., Pawlita, M. and Müller, M. (2013) *High-throughput pseudovirion-based neutralization assay for analysis of natural and vaccine-induced antibodies against human papillomaviruses* PLoS One, **8**: e75677

**Steele, J.**, Collins, S., Wen, K., Ryan, G., Constandinou-Williams, C. and Woodman, C.B.J. (2008) *Measurement of the humoral immune response following an incident human papillomavirus type 16 or 18 infection in young women by a pseudovirion-based neutralizing antibody assay* Clin. Vaccine Immunol., **15**, 1387-1390 (**B**)

### Neutralizing antibodies

#### Capsid protein

**Culp, T.D.**, Spatz, C.M., Reed, C.A. and Christensen, N.D. (2007) *Binding and neutralization efficiencies of monoclonal antibodies, Fab fragments and scFv specific for L1 epitopes on the capsid of infectious HPV particles* Virology, **361**, 435-446

#### L1 epitopes

**Carter, J.J.**, Wipf, G.C., Madelaine, M.M., Schwartz, S.M., Koutsky, L.A. and Galloway, D.A. (2006) *Identification of human papillomavirus type 16 L1 surface loops required for neutralization by human sera* J. Virol., **80**, 4664-4672

**Roth, S.D.**, Sapp, M., Streeck, R.E. and Selinka, H-C. (2006) *Characterization of neutralizing epitopes within the major capsid protein of human papillomavirus type 33* Virol. J., **3**:83

#### L2 epitopes:

**Kondo, K.**, Ishii, Y., Ochi, H., Matsumoto, T., Yoshikawa, H. and Kanda, T. (2007) *Neutralization of HPV16, 18, 31, and 58 pseudovirions with antisera induced by immunizing rabbits with synthetic peptides representing segments of the HPV16 minor capsid protein L2 surface region* Virology, **358**, 266-272

**Pastrana, D.V.**, Gambhira, R., Buck, C.B., Pang, Y-Y.S., Thompson, C.D., Culp, T.D., Christensen, N.D., Lowy, D.R., Schiller, J.T. and Roden, R.B.S. (2005) *Cross-neutralization of cutaneous and mucosal Papillomavirus types with anti-sea to the amino terminus of L2* Virology, **337**, 365-372 (**B, CTR**)

**Seitz, H.**, Schmitt, M., Böhmer, G., Kopp-Schneider, A. and Müller, M. (2013) *Natural variants in the major neutralizing epitope of human papillomavirus minor capsid protein L2* Int. J. Cancer, **132**, E139–E148

### Oropharyngeal cancer

**Conway, M.J.** and Meyers, C. (2009) *Replication and assembly of human papillomaviruses* Dent. Res., **88**, 307-317 (**CTR**)

## Ovarian cancer therapy

**Hung, C-F.**, Chiang, A.J., Tsai, H-H., Pomper, M.G., Kang, T.H., Roden, R.R. and Wu, T-C. (2012) *Ovarian cancer gene therapy using HPV-16 pseudovirion carrying the HSV-tk gene* PLoS One, **7**: e40983

**Kines, R.C.**, Cerio, R.J., Roberts, J.N., Thompson, C.D., de Los Pinos, E., Lowy, D.R. and Schiller, J.T. (2016) *Human papillomavirus capsids preferentially bind and infect tumor cells* Int. J. Cancer, **138**, 901–911

## Plants, production in

**Lamprecht, R.L.**, Kennedy, P., Huddy, S.M., Bethke, S., Hendrikse, M., Hitzeroth, I.I. and Rybicki, E.P. (2016) *Production of human papillomavirus pseudovirions in plants and their use in pseudovirion-based neutralization assays in mammalian cells* Sci. Rep., **6**: 20431

## Proteoglycans

**Huang, H-S.** and Lambert, P.F. (2012) *Use of an in vivo animal model for assessing the role of integrin  $\alpha_6\beta_4$  and Syndecan-1 in early steps in papillomavirus infection* Virology, **433**, 395–400

## Recombinant viruses

**Van Doorslaer, K.**, Porter, S., McKinney, C., Stepp, W.H. and McBride, A.A. (2016) *Novel recombinant papillomavirus genomes expressing selectable genes* Sci. Rep., **6**: 37782

## Serology

**Bissett, S.L.**, Wilkinson, D., Tettmar, K.I., Jones, N., Stanford, E., Panicker, G., Faust, H., Borrow, R., Soldan, K., Unger, E.R., Dillner, J., Minor, P. and Beddows, S. (2012) *Human papillomavirus antibody reference reagents for use in postvaccination surveillance serology* Clin. Vaccin. Immunol., **19** 449–451

**Dillner, J.** and Zhou, T. (2007) Meeting report of the WHO workshop and practical course on human papillomavirus (HPV) genotyping and HPV16/18 serology, Lusanne, Switzerland, June 2007

**Faust, H.**, Knekt, P., Forslund, O. and Dillner, J. (2010) *Validation of multiplexed human papillomavirus serology using pseudovirions bound to heparincoated beads* J. Gen. Virol., **91**, 1840–1848

## SNARE/Syntaxin

**Culp, T.D.**, Budgeon, L.R., Marinkovich, M.P., Meneguzzi, G. and Christensen, N.D. (2006) *Keratinocyte-secreted laminin 5 can function as a transient receptor for human papillomaviruses by binding virions and transferring them to adjacent cells* J. Virol., **80**, 8940-8950

**Laniosz, V.**, Nguyen, K.C. and Meneses, P.I. (2007) *Bovine papillomavirus type 1 infection is mediated by SNARE Syntaxin 18* J. Virol., **81**, 7435-7448 (B)

## Sublingual immunization

**Lee, H-J.**, Cho, H., Kim, M-G., Heo, Y-K., Cho, Y., Gwon, Y-D., Park, K.H., Jin, H., Kim, J., Oh, Y-K. and Kim, Y.B. (2015) *Sublingual immunization of trivalent human papillomavirus DNA vaccine in baculovirus nanovector for protection against vaginal challenge* PLoS One, **10**: e0119408

## Survival

**Ding, D-C.**, Chang, Y-C., Liu, H-W. and Chu, T-Y (2011) *Long-term persistence of human papillomavirus in environments* Gynecol. Oncol., **121**, 148–151

## Transfection

**Buck, C.B.**, Pastrana, D.V., Lowy, D.R. and Schiller, J.T (2005) *Generation of HPV pseudovirions using transfection and their use in neutralization assays* Methods Mol. Med., **119**, 445-462

## UV radiation

**Uberoi, A.**, Yoshida, S., Frazer, I.H., Pitot, H.C. and Lambert, P.F. (2016) *Role of ultraviolet radiation in papillomavirus-induced disease* PLoS Pathog., **12**: e1005664

## Vaccines/vaccination

### A7/A9 species groups

**Draper, E.**, Bissett, S.L., Howell-Jones, R., Edwards, D., Munslow, G., Soldan, K. and Beddows, S. (2011) *Neutralization of non-vaccine human papillomavirus pseudoviruses from the A7 and A9 species groups by bivalent HPV vaccine sera* Vaccine. **29**, 8585– 8590

### Antibody detection

Nie, J., Huang, W., Wu, X. and Wang, Y. (2014) *Optimization and validation of a high throughput method for detecting neutralizing antibodies against human papillomavirus (HPV) based on pseudovirions* J. Med. Virol., **86**, 1542–1555

### Antibody neutralization assay

Guan, J., Bywaters, S.M., Brendle, S.A., Lee, H., Ashley, R.E., Makhov, A.M., Conway, J.F., Christensen, N.D. and Hafenstein, S. (2015) *Structural comparison of four different antibodies interacting with human papillomavirus 16 and mechanisms of neutralization* Virology, **483**, 253–263

Panicker, G., Rajbhandari, I., Gurbaxani, B.M., Querec, T.D. and Unger, E.R. (2015) *Development and evaluation of multiplexed immunoassay for detection of antibodies to HPV vaccine types* J. Immunol. Methods, **417**, 107–114

Schellenbacher, C., Shafti-Keramat, S., Huber, B., Fink, D., Brandt, S. and Kimbauer, R. (2015) *Establishment of an in vitro equine papillomavirus type2 (EcPV2) neutralization assay and a VLP-based vaccine for protection of equids against EcPV2-associated genital tumors* Virology, **486**, 284–290 (E)

Sehr, P., Rubio, I., Seitz, H., Putzker, K., Ribeiro-Müller, L., Pawlita, M. and Müller, M. (2013) *High-throughput pseudovirion-based neutralization assay for analysis of natural and vaccine-induced antibodies against human papillomaviruses* PLoS One, **8**: e75677

### Calreticulin

Kim, D., Gambhira, R., Karanam, B., Monie, A., Hung, C-F., Roden, R. and Wu, T-C. (2008) *Generation and characterization of a preventative and therapeutic HPV DNA vaccine* Vaccine, **26**, 351-360

### Capsid variants

Bissett, S.L., Godi, A., Fleury, M.J.J., Touze, A., Cocuzza, C. and Beddows, S. (2015) *Naturally occurring capsid protein variants of human papillomavirus genotype 31 represent a single L1 serotype* J. Virol., **89**, 7748-7757

Mejia, A.F., Culp, T.D., Cladel, N.M., Balogh, K.K., Budgeon, L.R., Buck, C.B. and Christensen, N.D. (2006) *Preclinical Model To Test Human Papillomavirus Virus (HPV) Capsid vaccines in vivo using infectious HPV/cottontail rabbit papillomavirus chimeric papillomavirus particles* J. Virol., **80**, 12393-12397 (CTR)

### Capsomer vaccines

Wu, W-H., Gersch, E., Kwak, K., Jagu, S., Karanam, B., Huh, W.K., Garcea, R.L. and Roden, R.B.R. (2011) *Capsomer vaccines protect mice from vaginal challenge with human papillomavirus* PLoS One **6**: e27141

### DNA vaccines

Graham, B.S., Kines, R.C., Corbett, K.S., Nicewonger, J., Johnson, T.R., Chen, M., LaVigne, D., Roberts, J.N., Cuburu, N., Schiller, J.T. and Buck, C.B. (2010) *Mucosal delivery of human papillomavirus pseudovirus-encapsidated plasmids improves the potency of DNA vaccination* Mucosal Immunol., **5**, 475-486

Kines, R.C., Zarnitsyn, V., Johnson, T.R., Pang, Y-Y.S., Corbett, K.S., Nicewonger, J.D., Gangopadhyay, A., Chen, M. et al (2015) *Vaccination with human papillomavirus pseudovirus-encapsidated plasmids targeted to skin using microneedles* PLoS One, **10**: e0120797

Peng, S., Monie, A., Kang, T.H., Hung, C-F., Roden, R. and Wu, T-C. (2010) *Efficient delivery of DNA vaccines using human papillomavirus pseudovirions* Gene Ther., **17**, 1453–1464

Yang, B., Yang, A., Peng, S., Pang, X., Roden, R.B.S., Wu, T-C. and Hung, C-F. (2015) *Co-administration with DNA encoding papillomavirus capsid proteins enhances the antitumor effects generated by therapeutic HPV DNA vaccination* Cell Biosci., **5**:35

### E2 epitopes

Qian, J., Dong, Y., Pang, Y-Y.s., Ibrahim, R., Berzofsky, J.A., Schiller, J.T. and Kheif, S.N. (2006) *Combined prophylactic and therapeutic cancer vaccine: Enhancing CTL responses to HPV16 E2 using a chimeric VLP in HLA-A2 mice* Int. J. Cancer, **118**, 3022-3029

### Fusion protein

Karanam, B., Gambhira, R., Peng, S., Jagu, S., Kim, D.J., Ketner, G.W., Stern, P.L., Adams, R.J. and Roden, R.B.S. (2009) *Vaccination with HPV16 L2E6E7 fusion protein in GPI-0100 adjuvant elicits protective humoral and cell-mediated immunity* Vaccine **27**, 1040–1049

## Gardasil®

**Han, J.E.**, Kim, H.K., Park, S.A., Lee, S.J., Kim, H.J., Son, G.H., Kim, Y.T., Cho, Y.J., Kim, H-J. and Lee, N.G. (2010) *A nontoxic derivative of lipopolysaccharide increases immune responses to Gardasil® HPV vaccine in mice* Int. Immunopharmacol., **10**, 169-176

## Immunity and immune responses

**Giannini, S.L.**, Hanon, E., Moris, P., Van Mechelen, M., Morel, S., Dessy, F., Fourneau, M.A., Colau, B., Suzich, J., Losonsky, G., Martin, M-T., Dubin, G. and Wettendorf, M.A. (2006) *Enhanced humoral and memory B cellular immunity using HPV16/18 L1 VLP vaccine formulated with the MPL/aluminium salt combination (AS04) compared to aluminium salt only* Vaccine, **24**, 5937-5949

**Han, J.E.**, Kim, H.K., Park, S.A., Lee, S.J., Kim, H.J., Son, G.H., Kim, Y.T., Cho, Y.J., Kim, H-J. and Lee, N.G. (2010) *A nontoxic derivative of lipopolysaccharide increases immune responses to Gardasil® HPV vaccine in mice* Int. Immunopharmacol., **10**, 169-176

**Handisurya, A.**, Day, P.M., Thompson, C.D., Bonelli, M., Lowy, D.R. and Schiller, J.T. (2014) *Strain-specific properties and T cells regulate the susceptibility to papilloma induction by Mus musculus papillomavirus 1* PLoS Pathog., **10**: e1004314 (M)

**Hassett, K.J.**, Meinerz, N.M., Semmelmann, F., Cousins, M.C., Garcea, R.L. and Randolph, T.W. (2015) *Development of a highly thermostable, adjuvanted human papillomavirus vaccine* Eur. J. Pharm. Biopharm., **94**, 220–228

**Huo, Z.**, Bissett, S.L., Gienza, R., Beddows, S., Oeser, C. and Lewis, D.J.M. (2012) *Systemic and mucosal immune responses to sublingual or intramuscular human papilloma virus antigens in healthy female volunteers* PLoS One, **7**: e33736

**Kim, H.J.**, Lim, S.J., Kwag, H-L. and Kim, H-J. (2012) *The choice of resin-bound ligand affects the structure and immunogenicity of column-purified human papillomavirus type 16 virus-like particles* PLoS One **7**: e35893

**Peng, S.**, Ma, B., Chen, S-H., Hung, C-F. and Wu, T.C. (2011) *DNA vaccines delivered by human papillomavirus pseudovirions as a promising approach for generating antigen-specific CD8<sup>+</sup> T cell immunity* Cell Biosci., **1**: 26

**Tam, J.C.H.**, Bidgood, S.R., McEwan, W.A. and James, L.C. (2014) *Intracellular sensing of complement C3 activates cell autonomous immunity* Science, **345**: 1256070

**Vinzón, S.E.**, Braspenning-Wesch, I., Müller, M., Geissler, E.K., Nindl, I., Gröne, H-J., Schäfer, K. and Rösl, F. (2014) *Protective vaccination against papillomavirus-induced skin tumors under immuno-competent and immunosuppressive conditions: a preclinical study using a natural outbred animal model* PLoS Pathog., **10**: e1003924

## Immunoassay

**Panicker, G.**, Rajbhandari, I and Unger, E. (2012) *Detection of antibodies to HPV vaccine types using a multiplexed immunoassay* FASEB J., **26**, 577.8

**Panicker, G.**, Rajbhandari, I., Gurbaxani, B.M., Querec, T.D. and Unger, E.R. (2015) *Development and evaluation of multiplexed immunoassay for detection of antibodies to HPV vaccine types* J. Immunol. Methods, **417**, 107–114

## Immunogenicity enhancement

**Cho, H.**, Lee, H-J., Heo, Y-K., Cho, Y., Gwon, Y-D., Kim, M-G., Park, K.H., Oh, Y-K. and Kim, Y.B. (2014) *Immunogenicity of a trivalent human papillomavirus L1 DNA-encapsidated, non-replicable baculovirus nanovaccine* PLoS One, **9**: e95961

**Chang, D.Y.**, Kim, H.J. and Kim, H-J. (2012) *Effects of downstream processing on structural integrity and immunogenicity in the manufacture of papillomavirus type 16 L1 virus-like particles* Biotechnol. Bioproc. Eng., **17**, 755-763

**Lee, H-J.**, Hur, Y-K., Cho, Y-D., Kim, M-G., Lee, H-T., Oh, Y-K. and Kim, Y.B. (2012) *Immunogenicity of bivalent human papillomavirus DNA vaccine using human endogenous retrovirus envelope-coated baculoviral vectors in mice and pigs* PLoS One, **7**: e50296

**Nieto, K.**, Weghofer, M., Sehr, P., Ritter, M., Sedlmeier, S., Karanam, B., Seitz, H., Müller, M., Kellner, M., Hörer, M., Michaelis, U., Roden, R.B.S., Gissmann, L., Kleinschmidt, J.A. (2012) *Development of AAVLP(HPV16/31L2) particles as broadly protective HPV vaccine candidate* PLoS One, **7**: e39741

**Schellenbacher, C.**, Roden, R. and Kirnbauer, R. (2009) *Chimeric L1-L2 virus-like particles as potential broad-spectrum human papillomavirus vaccines* J. Virol., **83**, 10085-10095

### L1 antibodies

- Baud, D.**, Ponci, F., Bobst, M., De Gandhi, P. and Nardelli-Haeffliger, D. (2004) *Improved efficiency of a Salmonella-based vaccine against human papillomavirus type 16 virus-like particles achieved by using a codon-optimized version of L1* J. Virol., **78**, 12901-12909
- Berg, M.G.**, Adams, R.J., Gambhira, R., Siracusa, M.C., Scott, A.L., Roden, R.B.S. and Ketner, G. (2014) *Immune responses in macaques to a prototype recombinant adenovirus live oral human papillomavirus 16 vaccine* Clin. Vaccine Immunol., **21**, 1224–1231 (MQ)
- Bissett, S.L.**, Godi, A., Fleury, M.J.J., Touze, A., Cocuzza, C. and Beddows, S. (2015) *Naturally occurring capsid protein variants of human papillomavirus genotype 31 represent a single L1 serotype* J. Virol., **89**, 7748-7757
- Frailery, D.**, Zosso, N. and Nardelli-Haeffliger, D (2009) *Rectal and vaginal immunization of mice with human papillomavirus L1 virus-like particles* Vaccine, **27**. 2326–2334
- Qian, J.**, Dong, Y., Pang, Y-Y.s., Ibrahim, R., Berzofsky, J.A., Schiller, J.T. and Kheif, S.N. (2006) *Combined prophylactic and therapeutic cancer vaccine: Enhancing CTL responses to HPV16 E2 using a chimeric VLP in HLA-A2 mice* Int. J. Cancer, **118**, 3022-3029
- Rao, N.H.**, Babu, P.B., Rajendra, L., Sriraman, R., Pang, Y-Y.S., Schiller, J.T. and Srinivasan, V.A. (2011) *Expression of codon optimized major capsid protein (L1) of human papillomavirus type 16 and 18 in Pichia pastoris; purification and characterization of the virus-like particles* Vaccine, **29**, 7326– 7334
- Yang, B.**, Yang, A., Peng, S., Pang, X., Roden, R.B.S., Wu, T-C. and Hung, C-F. (2015) *Co-administration with DNA encoding papillomavirus capsid proteins enhances the antitumor effects generated by therapeutic HPV DNA vaccination* Cell Biosci., **5**:35

### L2 antibodies

- Conway, M.J.**, Cruz, L., Alam, S., Christensen, N.D., and Meyers, C. (2011) *Cross-neutralization potential of native human papillomavirus N-terminal L2 epitopes* PLoS One **6**: e16405
- Jagu, S.**, Malandro, N., Kwak, K., Yuan, H., Schlegel, R., Palmer, K.E., Huh, W.K., Campo, M.S. and Roden, R.B.S. (2011) *A multimeric L2 vaccine for prevention of animal papillomavirus infections* Virology, **420**, 43–50 (C, B)
- Jagu, S.**, Kwak, K., Schiller, J.T., Lowy, D.R., Kleanthous, H., Kalnin, K., Wang, C., Wang, H-K., Chow, L.T., Huh, W.K., Jaganathan, K.S., Chivukula, S.V. and Roden, R.B.S. (2013) *Phylogenetic considerations in designing a broadly protective multimeric L2 vaccine* J. Virol., **87**, 6127–6136
- Jagu, S.**, Karanam, B., Wang, J.W., Zayed, H., Weghofer, M., Brendle, S.A., Balogh, K.K., Tossi, K.P., Roden, R.B.S. and Christensen, N.D. (2015) *Durable immunity to oncogenic human papillomaviruses elicited by adjuvanted recombinant Adeno-associated virus-like particle immunogen displaying L2 17–36 epitopes* Vaccine **33** (2015) 5553–5563
- Kwak, K.**, Jiang, R., Wang, J.W., Jagu, S., Kirnbauer, R. and Roden, R.B.S. (2014) *Impact of inhibitors and L2 antibodies upon the infectivity of diverse alpha and beta human papillomavirus types* PLoS One, **9**: e97232
- Longet, S.**, Schiller, J.T., Bobst, M., Jichlinski, P. and Nardelli-Haeffliger, D. (2011) *A murine genital-challenge model is a sensitive measure of protective antibodies against human papillomavirus infection* J. Virol., **85**, 13253–13259
- Seitz, H.**, Schmitt, M., Böhmer, G., Kopp-Schneider, A. and Müller, M. (2013) *Natural variants in the major neutralizing epitope of human papillomavirus minor capsid protein L2* Int. J. Cancer, **132**, E139–E148
- Yang, B.**, Yang, A., Peng, S., Pang, X., Roden, R.B.S., Wu, T-C. and Hung, C-F. (2015) *Co-administration with DNA encoding papillomavirus capsid proteins enhances the antitumor effects generated by therapeutic HPV DNA vaccination* Cell Biosci., **5**:35
- Yoon, S-W.**, Lee, T-Y., Kim, S-J., Lee, I-H., Sung, M-H., Park, J-S. and Poo, H. (2012) *Oral administration of HPV-16 L2 displayed on Lactobacillus casei induces systematic and mucosal cross-neutralizing effects in Balb/c mice* Vaccine, **30**, 3286– 3294

### LPS-derivative (non-toxic)

- Han, J.E.**, Kim, H.K., Park, S.A., Lee, S.J., Kim, H.J., Son, G.H., Kim, Y.T., Cho, Y.J., Kim, H-J. and Lee, N.G. (2010) *A nontoxic derivative of lipopolysaccharide increases immune responses to Gardasil® HPV vaccine in mice* Int. Immunopharmacol., **10**, 169-176

### Measles-vectored vaccine

- Gupta, G.**, Giannino, V., Rishi, N. and Glueck, R. (2016) *Immunogenicity of next-generation HPV vaccines in non-human primates: Measles-vectored HPV vaccine versus Pichia pastoris recombinant protein vaccine* Vaccine, **34**, 4724–4731

### Memory B cells

**Scherer, E.M.**, Smith, R.A., Simonich, C.A., Niyonzima, N., Carter, J.J. and Galloway, D.A. (2014) *Characteristics of memory B cells elicited by a highly efficacious HPV vaccine in subjects with no pre-existing immunity* PLoS Pathog., **10**: e1004461

### MUC1 peptide – VLP conjugate

**Pejawar-Gaddy, S.**, Rajawat, Y., Hilioti, Z., Xue, J., Gaddy, D.F., Finn, O.J., Viscidi, R.P. and Bossis, I. (2010) *Generation of a tumor vaccine candidate based on conjugation of a MUC1 peptide to polyionic papillomavirus virus-like particles* Cancer Immunol. Immunother. **59**, 1685–1696 (B)

### Placental malarial antigen

**Thrane, S.**, Janitzek, C.M., Agerbæk, M.O., Ditlev, S.B., Resende, M., Nielsen, M.A., Theander, T.G., Salanti, A. and Sander, A.F. (2015) *A novel virus-like particle based vaccine platform displaying the placental malaria antigen VAR2CSA* PLoS One, **10**: e0143071

### Plants, virus production in

**Matić, S.**, Masenga, V., Poli, A., Rinaldi, R., Milne, R.G., Vecchiati, M. and Noris, E. (2012) *Comparative analysis of recombinant human papillomavirus 8 L1 production in plants by a variety of expression systems and purification methods* Plant Biotechnol. J., **10**, 410–421

### Salmonella-based

**Baud, D.**, Ponci, F., Bobst, M., De Gandhi, P. and Nardelli-Haeffliger, D. (2004) *Improved efficiency of a Salmonella-based vaccine against human papillomavirus type 16 virus-like particles achieved by using a codon-optimized version of L1* J. Virol., **78**, 12901-12909

**Frailery, D.**, Baud, D., Pang, S.Y-Y., Schiller, J., Bobst, M., Zosso, N., Ponci, F. and Nardelli-Haeffliger, D. (2007) *Salmonella enterica serovar typhi Ty21a expressing human papillomavirus type 16 L1 as a potential live vaccine against cervical cancer and typhoid fever* Clin. Vaccine Immunol., **14**, 1285-129

### Thermostability

**Hassett, K.J.**, Meinerz, N.M., Semmelmann, F., Cousins, M.C., Garcea, R.L. and Randolph, T.W. (2015) *Development of a highly thermostable, adjuvanted human papillomavirus vaccine* Eur. J. Pharm. Biopharm., **94**, 220–228

### Vaccination status

**Grant, B.D.**, Smith, C.A., Castle, P.E., Scheurer, M.E. and Richards-Kortum, R. (2016) *A paper-based immunoassay to determine HPV vaccination status at the point-of-care* Vaccine, **34**, 5656–5663

### VLP-vaccines

**Huber, B.**, Schellenbacher, C., Jindra, C., Fink, D. Shafti-Keramat, S. Kirnbauer, R. (2015) *A chimeric 18L1-45RG1 virus-like particle vaccine cross-protects against oncogenic alpha-7 human papillomavirus types* PLoS One, **10**: e0120152

### Yeast-expressed

**Kim, S.N.**, Jeong, H.S., Park, S.N. and Kim, H-J. (2007) *Purification and immunogenicity study of human papillomavirus type 16 L1 protein in Saccharomyces cerevisiae* J. Virol. Methods, **139**, 24-30

**Kwag, H-L.**, Kim, H.J., Chang, D.Y. and Kim, H-J. (2012) *The production and immunogenicity of human papillomavirus type 58 virus-like particles produced in Saccharomyces cerevisiae* J. Microbiol., **50**, 813-820

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